**Supplemental Material**

*Transmission Electron Microscopy (TEM)*

Clots were immersion-fixed overnight in 4% paraformaldehyde/0.15% glutaraldehyde in 0.15M sodium phosphate buffer (pH 7.4). Fixed clots were removed from the wells, washed, embedded in PolyBed 812 epoxy resin (Polysciences, Warrington, PA), and cross-sectioned (70 - 80 nm) to visualize fibrin. Clots were imaged using a LEO EM910 transmission electron microscope (Carl Zeiss SMT, Peabody, MA) operating at 80 kV. Digital images were recorded using a Gatan Orius CCD Camera and Digital Micrograph 3.11.0 (Gatan INC, Pleasanton, CA).

*Gel Electrophoresis*

Sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) was performed to detect the constituents of the purified clot samples. Fibrin clot samples (150 µL) were prepared with the same procedures described in the methods section. Three hours after preparation into 2 mL tubes clot samples were centrifuged at 5000×*g* for 20 minutes. Supernatant (50 µL) from each sample was mixed with 50 µL 2X SDS sample buffer. The solution was boiled for 5 min and then frozen at -75 °C. Upon thawing, samples were boiled again for 5 min and proteins were separated by SDS-PAGE on a Phastgel System (Pharmacia Biotech) according to the manufacturer’s instructions.

*Statistical Methods*

Statistical analyses were adjusted for multiple sample testing using a Bonferroni correction and a linear mixed model was implemented with *post hoc* comparison between blood samples using the normal blood as the reference. Statistical analysis was carried out using IBM SPSS 20 (SPSS Inc., Chicago, IL). We considered a *p*-value smaller than 0.05 as statistically significant.